

The Structures of Amphidinolide B Isomers: Strongly Cytotoxic Macrolides Produced by a Free-Swimming Dinoflagellate, *Amphidinium* sp.

Ines Bauer, Lucie Maranda, and Yuzuru Shimizu*

Department of Pharmacognosy and Environmental Health Sciences, College of Pharmacy
The University of Rhode Island
Kingston, Rhode Island 02881

Russell W. Peterson and Laurie Cornell

Bristol-Myers Squibb Pharmaceutical Research Institute
Department of Experimental Therapeutics, P.O. Box 4000
Princeton, New Jersey 08543

Jorge Rios Steiner and Jon Clardy*

Department of Chemistry, Baker Laboratory
Cornell University, Ithaca, New York 14853-1301

Received December 8, 1993

Interesting macrolides have been isolated from dinoflagellates, genus *Amphidinium*, symbiotic with the marine flatworms, *Amphiscolops* spp. Many of them have shown potent toxicity against tumor cell lines and are attracting attention as potential cancer drugs.¹⁻¹⁰ The planar structures of these dinoflagellate macrolides have been elucidated primarily by means of 2D-NMR, but their stereochemistry remains unsolved except for that of the small simple macrolide amphidinolide J.⁵ In this communication we report the establishment of the relative stereochemistry of 26-membered C₃₂ macrolides in the amphidinolide B group, the most representative and important group of amphidinolides.

The free-swimming dinoflagellate, strain S1-36-5,¹¹ was collected at Brewers Beach, St. Thomas, U.S. Virgin Islands, cloned, and cultured in seawater enriched with K-supplements.¹² From the freeze-dried algal cells, three isomeric compounds, which we tentatively call amphidinolides B₁, B₂, and B₃ in this paper, were isolated in yields of 0.14%, 0.024%, and 0.0076% of the dried cell weight, respectively.¹³

(1) Kobayashi, J.; Ishibashi, M.; Nakamura, H.; Ohizumi, Y.; Yamasu, T.; Sasaki, T.; Hirata, Y. *Tetrahedron Lett.* 1986, 47, 5755-5758.

(2) Ishibashi, M.; Ohizumi, Y.; Hamashima, M.; Nakamura, H.; Hirata, Y.; Sasaki, T.; Kobayashi, J. *J. Chem. Soc., Chem. Commun.* 1987, 1127.

(3) Kobayashi, J.; Ishibashi, M.; Wälchli, M. R.; Nakamura, H.; Hirata, Y.; Sasaki, T.; Ohizumi, Y. *J. Am. Chem. Soc.* 1988, 110, 490-494.

(4) Kobayashi, J.; Ishibashi, M.; Nakamura, H.; Ohizumi, Y.; Hirata, Y.; Sasaki, T.; Ohta, T.; Nozoe, S. *J. Nat. Prod.* 1989, 52, 1036-1041.

(5) Kobayashi, J.; Sato, M.; Ishibashi, M. *J. Org. Chem.* 1993, 58, 2645-2646.

(6) Kobayashi, J.; Ishibashi, M.; Murayama, T.; Takamatsu, M.; Iwamura, M.; Ohizumi, Y.; Sasaki, T. *J. Org. Chem.* 1990, 55, 3421-3425.

(7) Ishibashi, M.; Sato, M.; Kobayashi, J. *J. Org. Chem.* 1993, 58, 6928-6929.

(8) Kobayashi, J.; Tsuda, M.; Ishibashi, M.; Shigemori, H.; Yamasu, T.; Hirota, H.; Sasaki, T. *J. Antibiot.* 1991, 44, 1259-1261.

(9) Kobayashi, J.; Shigemori, H.; Ishibashi, M.; Yamasu, T.; Hirota, H.; Sasaki, T. *J. Org. Chem.* 1991, 56, 5221-5224.

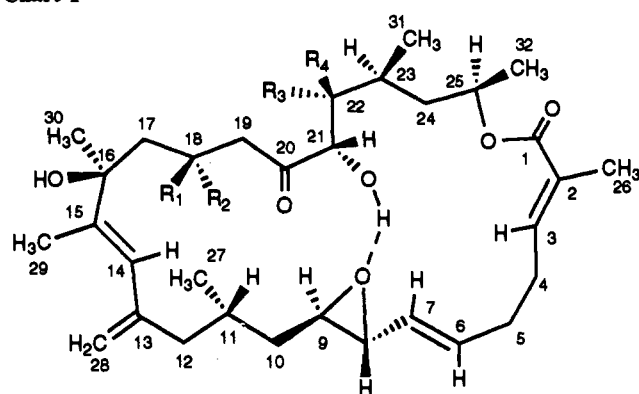
(10) Kobayashi, J.; Ishibashi, M. *Chem. Rev.* 1993, 93, 1753-1770.

(11) Compared to most other *Amphidinium* species, the organism is very large in size (length, 31-43 μ m; width, 19-23 μ m). Although it best fits the description of *A. carterae* Hulbert, no definite identification has been made. Morphological and ultrastructural studies are under way and will be published elsewhere.

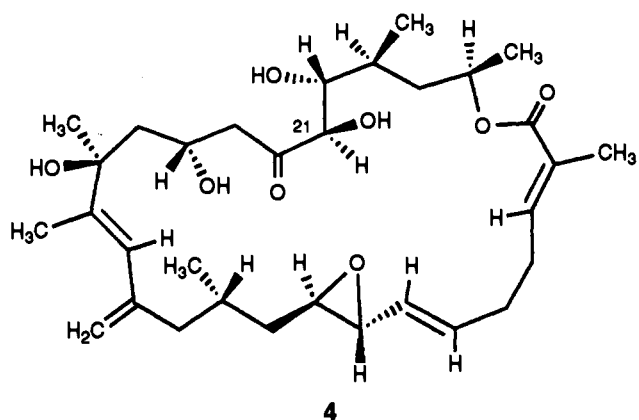
(12) Keller, M. D.; Selvin, R. C.; Claus, W.; Guillard, R. R. L. *J. Phycol.* 1987, 23, 633-638.

(13) The toluene-methanol extract of the cells was partitioned between *n*-hexane and 90% MeOH. The methanol layer was chromatographed successively on silica gel (CH₂Cl₂/MeOH, 97:3), C18 silica gel (CH₃CN/H₂O, 8:2), and Hamilton PRP-1 (CH₃CN/H₂O, 8:2) and CN silica gels (Isooctane/2-PrOH, 8:1) to afford pure compounds.

Chart 1



1. R₁, R₄=H; R₂, R₃=OH
2. R₁, R₃=OH; R₂, R₄=H
3. R₂, R₄=OH; R₁, R₃=H



4

Amphidinolide B₁ (1, Chart 1) was obtained as needles from *n*-hexane-CH₂Cl₂, mp 82-84 °C; [α]_D²⁵ = -62.5 \pm 0.5° (*c* 0.39, CHCl₃), [α]_D²⁵ = -71.3 \pm 0.4° (*c* 0.39, CH₂Cl₂); HRFABMS *m/z* 563.359 680 (C₃₂H₅₀O₈, MH⁺, Δ -2.3 ppm); UV λ_{\max} = 220 nm (MeOH, ϵ 17 000). The NMR spectral data of 1 were almost identical with those reported for amphidinolide B by Ishibashi *et al.*² The structure deduced from the 2D-NMR spectra was also in agreement with the revised planar structure of amphidinolide B by Kobayashi *et al.*⁴

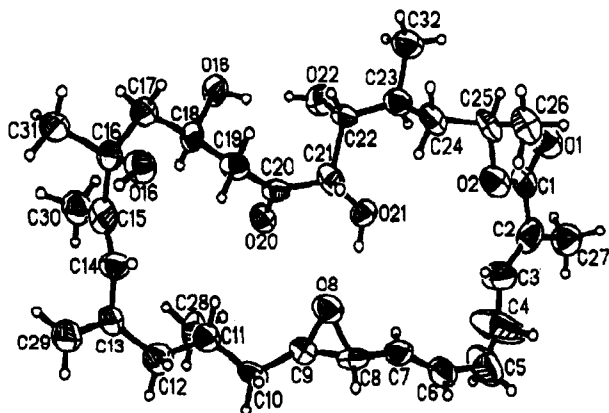
The relative stereochemistry of 1 was obtained from a single crystal X-ray diffraction analysis.¹⁴ Figure 1 shows a perspective view of the final X-ray model. Amphidinolide B₁ has a rectangular shape, which is bridged in the middle by a hydrogen bond (2.02 Å) between 21-OH and the epoxide O. The solution conformation of 1 in CHCl₃ and CH₂Cl₂ seems to be close to the

(14) A single crystal of 1 (0.15 \times 0.35 \times 0.50 mm³) was covered with a thin layer of paratone N oil and mounted in a capillary. Data were collected at -40 °C on a Siemens R3M diffractometer to $2\theta \leq 115^\circ$ (Cu K α) using variable speed ω scans from 2.50 to 29.30 deg/min. Accurate cell constants, determined by a least-squares fit of 25 well-centered reflections ($25^\circ \leq 2\theta \leq 30^\circ$), were $a = 11.417(7)$, $b = 15.220(4)$, and $c = 22.486(6)$ Å. Systematic extinctions uniquely indicated space group $P2_12_12_1$ with one molecule of C₃₂H₅₀O₈ plus a C₆H₁₂ solvent constituting the asymmetric unit. Periodically monitored check reflections showed no significant decay. A total of 2991 unique reflections were collected, of which only 1646 (55%) were considered observed ($|F| > 3.0\sigma(F)$). Reflections were corrected for Lorentz and polarization but not absorption. The structure was solved by direct methods using the Siemens SHELXTL PLUS library. Blocked full-matrix least-squares refinements for all positional and thermal parameters for all non-hydrogen atoms and isotropic thermal parameters with partial occupancies for most of the solvent atoms were used. Hydrogens were included using the riding model. Refinement indicates significant thermal disorder for the solvent molecule and serious librational motion within the C3 to C6 region (C4-C5 bond is 1.137 Å, due to this effect). A total of 401 parameters were used in the refinements, to give a final unweighted $R = 9.90\%$ and $R_w = 13.62\%$, with $w^{-1} = \sigma^2(F) + 0.0066F^2$. Additional data are available and described in the supplementary material.

Table 1. Comparison of ^1H NMR Chemical Shifts and Coupling Constants of H17–H23 of Amphidinolides B₁ (1), B₂ (2), and B₃ (3) and Coupling Constants Calculated from the Crystal Structure of 1

position	^1H NMR: δ (ppm) and J (Hz) obsd			J (Hz) calcd from the crystal structure of 1 ^a	
	1 (CDCl ₃)	2 (CDCl ₃)	3 (CDCl ₃)	1	structures inverted at C18 (*) and C22 (**)
17b	1.76 dd $J_{\text{H17b,18}} = 5.3$	1.79 dd $J_{\text{H17b,18}} = 7.6$	1.75 dd $J_{\text{H17b,18}} = 5.9$	$J_{\text{H17b,18}} = 2.6$	$J_{\text{H17b,18}} = 4.0^*$
18	4.16 m	4.19 m	4.16 m		
19a	2.87 dd $J_{\text{H19a,18}} = 7.2$	3.31 dd $J_{\text{H19a,18}} = 2.7$	3.03 dd $J_{\text{H19a,18}} = 8.3$	$J_{\text{H19a,18}} = 8.0$	$J_{\text{H19a,18}} = 2.5^*$
19b	2.78 dd $J_{\text{H19b,18}} = 3.3$	2.63 dd $J_{\text{H19b,18}} = 8.6$	2.87 dd $J_{\text{H19b,18}} = 3.5$	$J_{\text{H19b,18}} = 1.7$	$J_{\text{H19b,18}} = 8.0^*$
21	4.33 dd $J_{\text{H21,22}} = 1.9$	4.20 dd $J_{\text{H21,22}} = 2.0$	4.15 dd $J_{\text{H21,22}} = 8.4$	$J_{\text{H21,22}} = 3.4$	$J_{\text{H21,22}} = 7.4^{**}$
22	3.71 ddd $J_{\text{H21,22}} = 1.9$ $J_{\text{H22,23}} = 10.1$	3.65 ddd $J_{\text{H21,22}} = 2.0$ $J_{\text{H22,23}} = 10.8$	3.55 ddd $J_{\text{H21,22}} = 8.4$ $J_{\text{H22,23}} = 3.1$	$J_{\text{H22,21}} = 3.4$ $J_{\text{H22,23}} = 9.9$	$J_{\text{H22,21}} = 7.4^{**}$ $J_{\text{H22,23}} = 2.7^{**}$

^a Calculated according to the modified Karplus equation (Bothner-By, A. B. *Adv. Magn. Reson.* 1965, 1, 195) and corrected for the electron negativity of the substituents.

**Figure 1.** Perspective view of X-ray crystallographically-determined structure of amphidinolide B₁ (1).

crystal conformation, because the observed spin–spin coupling constants match the values calculated from dihedral angles obtained from the crystal structure.

Both amphidinolide B₂ (2), an amorphous solid, $[\alpha]_D^{25} = -43.9 \pm 0.2^\circ$ (*c* 0.42, CHCl₃); HRFABMS *m/z* 563.359 680 (C₃₂H₅₀O₈, MH⁺, $\Delta -2.3$ ppm); UV: $\lambda_{\text{max}} = 220$ nm (ϵ 16 000, MeOH), and amphidinolide B₃ (3), $[\alpha]_D^{25} = -69.4 \pm 1.1^\circ$ (*c* 0.16, CHCl₃); HRFABMS *m/z* 563.359 680 (C₃₂H₅₀O₈, MH⁺, $\Delta -2.3$ ppm); $\lambda_{\text{max}} = 220$ nm (ϵ 16 000, MeOH), showed NMR spectra very similar to those of 1. In the spectra of 2, the only significant differences are in the chemical shifts and coupling constants of H17 and H19 (Table 1). If 2 has the same conformation as 1, as is likely in view of their chemical shifts and coupling constants of the rest of the molecule, these differences can be best explained if 2 is the C18 stereoisomer of 1. Similarly, differences in the NMR spectra of 3 are confined around C22. There are differences in the coupling constants and chemical shifts of H22, and $J_{\text{H22,H23}}$ and $J_{\text{H22,H21}}$ are close to the calculated values for the C22 epimer (Table 1). Formulating 3 as the 22 epimer also explains the

downfield shifts observed for the C19 methylene hydrogens, because, in the 22 epimeric form, the hydroxyl group comes closer (~ 3 Å) to C19 methylene hydrogens.

Comparison of the ^1H and ^{13}C NMR data strongly indicates that amphidinolide B₁ (1) and amphidinolide B₂ (2) are identical with amphidinolide B² and amphidinolide D⁴, respectively. However, Kobayashi *et al.* assigned the structure of 21-*epi*-amphidinolide B (4) to amphidinolide D on the basis that amphidinolide D showed prominent NOEs between H21 and H19, as well as between H21 and 23-methyl protons. In amphidinolide B, such NOEs were absent.⁴ If the compounds are identical with 1 and 2, this result cannot be predicted on the basis of the crystal conformation. In the X-ray structure of 1, H21 is located very close to both H19a,b, and a strong NOE is expected. The stereochemistry of the 23-methyl group is such that no NOE seems possible with H21 in either 21 epimer. Additional experiments as well as caution will be needed to secure the unequivocal identities of these isomeric compounds.

Amphidinolides B₁, B₂, and B₃ exhibited potent cytotoxicity against human colon tumor cell line HCT 116 (IC₅₀ 0.122, 7.5, and 0.206 $\mu\text{g}/\text{mL}$, respectively). However, the level of the activity was significantly lower than that reported for amphidinolide B and amphidinolide D against a different cell line, murine leukemia L1210 (IC₅₀ 1.4×10^{-4} and 1.9×10^{-2} $\mu\text{g}/\text{mL}$, respectively).^{2,4}

Acknowledgment. We thank Dr. Michael A. McGregor for NMR measurements. This research was supported by NIH Grants CA 49992 (Y.S.) and CA 50750 (J.C. and Y.S.), which are greatly appreciated.

Supplementary Material Available: ^1H and ^{13}C NMR spectra and tables containing assignments and coupling constants of the compounds 1, 2 and 3, and X-ray data for 1 (37 pages). This material is contained in many libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.